Medical Progress

Acute Pancreatitis

MICHAEL C. GEOKAS, M.D., PH.D., Los Angeles

■ For many decades two types of acute pancreatitis have been recognized: the edematous or interstitial and the hemorrhagic or necrotic. In most cases acute pancreatitis is associated with alcoholism or biliary tract disease. Elevated serum or urinary α -amylase is the most important finding in diagnosis. The presence of methemal-bumin in serum and in peritoneal or pleural fluid supports the diagnosis of the hemorrhagic form of the disease in patients with a history and enzyme studies suggestive of pancreatitis.

There is no characteristic clinical picture in acute pancreatitis, and its complications are legion. Pancreatic pseudocyst is probably the most common and pancreatic abscess is the most serious complication.

The pathogenetic principle is autodigestion, but the precise sequence of biochemical events is unclear, especially the mode of trypsinogen activation and the role of lysosomal hydrolases. A host of metabolic derangements have been identified in acute pancreatitis, involving lipid, glucose, calcium and magnesium metabolism and changes of the blood clotting mechanism, to name but a few.

Medical treatment includes intestinal decompression, analysics, correction of hypovolemia and other supportive and protective measures. Surgical exploration is advisable in selected cases, when the diagnosis is in doubt, and is considered imperative in the presence of certain complications, especially pancreatic abscess.

"It is scant modesty for man, even if he is 'the highest vertebrate' to presume that he can predict the cosmic plan on the intensity of his joy and pain, or cement the stars together with even his highest aspirations."

HOMER SMITH. From Fish to Philosopher; Boston; Little, Brown, 1953.

THE PANCREATIC ACINAR CELL segregates the enzyme protein from the cytoplasm immediately after its synthesis in order to avoid autodigestion. The endoplasmic reticulum, the Golgi complex, the zymogen granule membrane and the cell membrane proper constitute an effective interposition device covering the entire secretory cycle of the acinar cell. The pancreatic proteases (trypsin, chymotrypsin, elastase and carboxypeptidases) and phospholipase A are stored in the acinar cell and exist in the pancreatic secretion,

From the Departments of Medicine, Sepulveda Veterans Administration Hospital and University of California, Los Angeles, School of Medicine.

Reprint requests to: M. C. Geokas, M.D., Department of Medicine, Sepulveda Veterans Administration Hospital, 16111 Plummer Street, Sepulveda, Ca. 91343.

before it enters the duodenum, as inactive zymogen.^{1,2} Trypsin plays the central role in the physiological activation of zymogen because the activation process consists of one or more trypsin catalyzed partial proteolysis reactions.³ The stability of the entire zymogen system largely depends on the trypsin inhibitors which under normal conditions will prevent the trypsin activation of zymogen in pancreatic tissue and pancreatic secretion into the pancreatic ductal system.⁴ Finally, healthy respiring acinar cells are not amenable to digestion by pancreatic proteases, because it has been shown that normal cell metabolism and cell membrane structure constitute important safeguards against enzyme-digestion.⁵

In acute pancreatitis inappropriate activation and release of enzymatic activity occurs in and around the pancreas^{6,7} in pancreatic juice,⁸ and into the blood stream.9 The disease has been induced in various animals such as dogs, rats, cats, rabbits, goats and monkeys by a variety of methods, and certain forms of experimental pancreatitis bear some resemblance in microscopic pathologic features and in biochemical changes of the disease in man. Since the brilliant experiment of Claude Bernard in 1856,10 two important questions remain unanswered, (1) how the various etiological factors initiate acute pancreatitis by overcoming the existing natural safeguards and (2) which mechanisms are responsible for the propagation of hydrolytic cleavage of cellular structures leading to generalized pancreatic necrosis and death.

Nonetheless, recent experimental and clinical data have provided some clues in the pathophysiology of acute pancreatitis and the time appears ripe to review previous information in the light of certain new developments. Since this review is mainly clinical, and recent reviews of etiological and pathogenetic mechanisms are available, 11,12 many important contributions will not be included. On the other hand, speculation and personal opinion on controversial points concerning pathophysiology, differential diagnosis, metabolic consequences and treatment will be discussed.

Etiology Alcoholic Pancreatitis

The mechanism by which the heavy drinking of alcohol leads to acute attacks of pancreatitis

remains unclear. Shapiro et al¹³ have proposed a detailed hypothesis which includes stimulation of gastric acid secretion both by gastrin release in the antral mucosa and by direct stimulation of parietal cells. In turn, acid reaching the duodenum would promote secretion of secretin and pancreozymin with resultant stimulation of pancreatic secretion by all three gastrointestinal hormones. Concomitantly, a direct effect of ethanol on the sphincter of Oddi would increase sphincteric resistance14-16 with resultant increase in intrapancreatic ductal pressure. Another proposed mechanism for alcoholic pancreatitis involves the reflux of bile into the pancreatic duct, but this suggestion remains at present purely hypothetical.¹⁷ On the other hand, sphincteroplasty has been shown to reduce the incidence of pancreatitis even in patients who continue the intake of ethanol following this procedure,18 a fact which probably indicates that reflux of bile does not play an important role in initiating alcoholic pancreatitis.

It has been shown that total obstruction of the pancreatic duct in dogs, combined with stimulation of pancreatic flow by endogenous or exogenous hormones, produces little pancreatic inflammatory response.19 Recent studies suggest that a direct toxic effect of ethanol on the pancreas should be considered as at least partially involved in the production of pancreatitis. In the liver cells, structural changes have been described which probably represent a direct hepatotoxic effect of ethanol,20-22 and it seems that a similarity exists between hepatic and pancreatic ultrastructural lesions, following the ingestion of ethanol in experimental animals. Darle et al²³ showed that in rats following long-term ethanol ingestion, the pancreas exhibited lipid droplets in acinar cells, swelling of mitochondria and reduction of their inner membranes, and cytoplasmic degradation of acinar and centroacinar and duct cells. Long-term ethanol ingestion is accompanied by decreased protein synthesis in the pancreas,24 and by inhibition of uptake of 32P into pancreatic phospholipids.25 Furthermore, impairment of Na+ dependent uptake of certain amino acids by pancreatic slices of alcoholtreated mice in vitro has been reported.26 Further studies are required, however, to substantiate the direct toxic effect of ethanol on the pancreatic acinar cells and the mode of production of alcoholic pancreatitis.

Hyperlipidemia

Hyperlipidemia at times seen with acute pancreatitis may be (a) primary and a cause of pancreatitis, (b) secondary and a consequence of pancreatitis, and (c) a casual association with acute pancreatitis. Two of the primary hyperlipoproteinemias (type I and V) may be associated with bouts of abdominal pain due to pancreatitis.²⁷ Attacks of pancreatitis frequently begin in childhood and over the years lipemia retinalis, hepatosplenomegaly, and any of the many forms of xanthomas may be observed. Accurate classification of the lipid abnormality can usually be obtained with lipoprotein electrophoresis.²⁸

Determination of lipoprotein lipase activity²⁹ is helpful in differentiating between type I and V. The cause of pancreatitis in familial hyperlipidemia is unknown. The episodes of acute pancreatitis are frequently mild and without severe complications.

Hypercalcemia

Hypercalcemia due to parathyroid adenoma,30 parathyroid carcinoma³¹ excessive doses of vitamin D,³² multiple myeloma or calcium carbonate antacid overdose³³ might result in acute pancreatitis. The relationship between hypercalcemia and pancreatitis remains an uncertain one. Regardless of cause, hypercalcemia leads to an increase in calcium concentration and content in pancreatic juice. Data on experiments with dogs have shown that a rise in volume and bicarbonate concentration in pancreatic secretion is not accompanied by an increase in calcium concentration. 34,35 In patients with hyperparathyroidism there is increased calcium concentration in basal pancreatic secretion³⁶ and the same is true in experimental hypercalcemia in animals.37 In chronic pancreatitis not related to hyperparathyroidism or hypercalcemia, there is high calcium content in basal pancreatic secretion. 36,38,39 It is noteworthy that the injection of soluble calcium compounds into the pancreatic ducts of rats results in severe acute pancreatitis.40 In hyperparathyroidism acute pancreatitis has been attributed to conversion of trypsinogen into trypsin by high levels of calcium in pancreatic tissue and pancreatic secretion. Thus, in hypercalcemic rats increased levels of trypsin were found in pancreatic juice,³⁷ but no similar observation has been made in man. Pancreatic calcification is

common in pancreatitis associated with alcoholism^{41,42} hyperparathyroidism,^{43,44} hereditary pancreatitis,^{45,46} pancreatitis secondary to a duct obstructing pancreatic carcinoma,⁴³ rarely in pancreatitis accompanying biliary tract disease⁴¹ and quite frequently in painless pancreatitis.

Solitary calculi obstructing the main pancreatic duct represent a variant of painless pancreatitis. The predominant form of calcium salt precipitation in the pancreas is calcium carbonate into the ducts, and less common is hydroxyapatite calcification deposited in areas of fat necrosis in the parenchyma.⁴⁷

Biliary Tract Disease

The causative factors involved in the association of biliary tract disease and acute pancreatitis remain obscure. Recent work has focused on two areas of investigation: (a) the reflux into the pancreatic duct of altered components of bile such as unconjugated bile salts⁴⁸ and lysolecithin,⁴⁹ and (b) the identification of lymphatic channels as a pathway for the propagation of toxins with resultant pancreatic inflammation.⁵⁰ Deconjugation of conjugated bile salts could be effected by certain bacteria⁴⁸ and lysolecithin could be produced by the phospholipase A of pancreatic secretion acting upon bile lecithin.⁴⁹

In one study unconjugated bile salts were detected in bile of patients with acute pancreatitis associated with gall bladder disease; and in another study,⁵¹ of patients with acute cholecystitis, lysolecithin was identified in gall bladder as well as in hepatic bile. However, the most common known organism in bile, in biliary tree disease, is E. coli⁵² which is unable to deconjugate bile salts. On the other hand, E. coli infected bile introduced under low pressure in the pancreatic ducts of cats had pronounced mucolytic and cytotoxic effects.⁵³

Weiner and his associates⁵⁰ have shown that in dogs lymphatic communications do exist between the biliary tract and the pancreas. When acute cholecystitis was produced, by staphylococcal toxin injected into the gall bladder lymphatics or by lipase injected into the gall bladder, acute pancreatitis developed in a high proportion of animals. Furthermore, when india ink was infused into the lymphatics of the gall bladder, it entered the lymphatics in the interlobular spaces of the pancreas. Conversely, india

ink infused into the pancreatic lymphatics by ductal injection was demonstrated in lymphatics along the common bile duct.

Dixon and Hillam⁵⁴ pointed out that any speculation concerning the mechanism of acute pancreatitis associated with biliary tract disease should explain pancreatitis occuring with cholesterosis of the gall bladder, acalculous cholecystitis, acute and chronic calculous cholecystitis and choledocholithiasis with and without a "common channel." These investigations advanced the following three interesting possibilities: (a) a common underlying factor may be present in biliary tract disease and pancreatitis; (b) biliary tree disease may cause pancreatitis and (c) pancreatitis may cause biliary tree disease.

It is obvious from the preceding considerations that additional studies are required in order to identify the intricate mechanisms involved in this association, such as the role of bile salt deconjugating bacteria—for example, bacteroides, clostridia and streptococcus fecalis. It is possible that bacterial toxins from the biliary tree could find access to the pancreas through anastomotic lymphatic channels and could produce pancreatitis by labilizing cellular membranes with resultant release of pancreatic hydrolases, intracellular as well as exportable.

Hereditary Pancreatitis

Hereditary pancreatitis is a rare condition that manifests itself in childhood, in about half of the patients, and is inherited as a non-sex-linked Mendelian dominant, but poor penetrance of the gene and incomplete recessiveness have not been excluded. All patients have been white and most of them of northern European ancestry. In their patients with familial pancreatitis, Gross and his associates⁵⁵ found daily excretions as high as 1,600 mg of lysine and 848 mg of cystine and suggested that this specific aminoaciduria might represent a genetic defect or some other inborn metabolic error. However, aminoaciduria might be seen in some patients with non-familial pancreatitis and might be absent in half of the patients with the familial form. In the latter case the disease should be suspected when blood relatives have similar attacks of abdominal pain, in the absence of biliary tract disease or alcoholism and when there is a history of attacks beginning in childhood. In about half of the reported cases calcification of the pancreas has been found to occur in the larger pancreatic ducts^{58,57} as gross calculi. Overt diabetes mellitus and exocrine insufficiency have been found in 20 to 25 percent of cases. The nature of the inherited predisposing abnormality remains unknown, and to date no other defect common to all affected persons has been recognized. Carcinoma of the pancreas has been reported to occur in some cases of hereditary pancreatitis.⁵⁸

Postoperative Pancreatitis

Acute pancreatitis is a well recognized postoperative complication, mainly after biliary and gastric surgical operation although at times it occurs after operations in areas remote from the pancreas-for example, thyroidectomy or transurethral prostatectomy or orthopedic operations.⁵⁹ Contributing etiologic factors are trauma to the pancreatic tissue and ducts, and duct obstruction or compromise of blood supply producing ischemia with resultant autodigestion. In instances where local trauma cannot be responsible, hypovolemic shock,60 the formation of microthrombi and a decrease of trypsin inhibitor in pancreatic juice have been cited as predisposing factors. The highest reported mortality in postoperative pancreatitis is 74 percent.⁶¹ In a series of cases of pancreatitis following biliary tract operations, reported by Bardenheier and his associates,62 common bile duct exploration and previous history of pancreatitis were considered the main contributory factors. The early postoperative diagnosis of pancreatitis is frequently difficult, and hypotension, oliguria, jaundice or a palpable mass in the upper abdomen, coupled with elevated serum and urinary amylase, should be helpful. With operative injury to the pancreas the amount of amylase in the urine increases, according to Ambromovage et al,63 and the increment appears to be related more to the functional state of the gland than to the magnitude of pancreatic injury. Keighley and his associates⁶⁴ indicated that damage to the sphincter of Oddi in traumatic explorations of the common bile duct and sphincterotomy appear to be the most important factors for postoperative hyperamylasemia and pancreatitis.

Several recent reports of acute pancreatitis following extracorporeal circulation have appeared. 65-67 Johnson and Nabseth reported a case of hemorrhagic pancreatitis following a cadaver renal transplant. Their survey of 1,321

renal transplants showed 23 cases of pancreatitis with 12 deaths. Speculated etiologic factors have been corticosteroid therapy, surgical trauma, induced auto-immune pancreatic rejection, decreased host protective responses and infection.

Pancreatitis in Pregnancy

The reason for the known tendency of acute pancreatitis of pregnancy to develop mostly during the last trimester or in the early postpartum period is obscure. Biochemical, hormonal and mechanical changes occurring during pregnancy and early puerperium might be important. To date, 106 cases of acute pancreatitis have been reported in association with pregnancy. 69 Biliary tract disease has been found in a substantial number of the cases. 70,71 Fatal hemorrhagic pancreatitis in pregnancy attributed to chlorothiazide administration has been reported,73 and hyperparathyroidism in pregnancy has also been known to be a contributing factor. Maternal hyperparathyroidism leading to suppression of fetal parathyroid glands with resultant tetany in the child is known to occur. 73 Maternal mortality of 24.3 percent in 37 cases observed since 1951 was found by Berk et al.69 Fetal mortality has also been high. Culdocentesis may be of special diagnostic value in pregnant women. The treatment is generally conservative, with termination of pregnancy to be considered when response to therapy is not satisfactory.

Drug-induced Pancreatitis

New cases of acute pancreatitis have been reported developing in the course of treatment with various therapeutic agents such as corticosteroids,74,75 agents producing necrotizing angiitis in various organs including the pancreas, 76. salicylazosulfapyridine⁷⁷ and thiazides.⁷⁸ Acute pancreatitis in patients with acute leukemia and lymphoma during treatment with L-asparaginase (obtained from both A and B strains of E. coli), has been recorded. 79,80 L-asparaginase is known to arrest DNA synthesis and mitosis, in regenerating rat liver and abnormal liver function tests are common during L-asparaginase therapy in man. The mechanism of this kind of acute pancreatitis in man, however, has not been elucidated.

Generalized Infections

Acute pancreatitis has been associated with inflamatory processes through (a) lymphatic propagation from adjacent structures such as the gall bladder, (b) direct extension as in peritonitis, and (c) diseases in which another mechanism, possibly hematogenous spread, has to be considered. Examples of acute pancreatitis occurring in the course of severe bacterial infections have been recorded in typhoid fever, salmonella typhimurium infection, s1 scarlet fever, s2 streptococcal food poisoning, s3 and dysentery. s4 Furthermore, viral infections known to cause pancreatitis in man include viral hepatitis, s5 infectious mononucleosis and mumps. Acute pancreatitis has also been observed in animals infected with the encephalomyocarditis, s8 coxsackie group B89 and foot and mouth disease viruses.

Miscellaneous

Acute pancreatitis has been found to be associated with a multiplicity of other conditions such as systemic lupus erythematosus, electric shock, 91 methyl alcohol poisoning and atheromatous embolization to the intrapancreatic arteries, 92 to name but a few. In Trinidad, the most common cause of acute pancreatitis seems to be the sting of a scorpion. 93 Of particular interest is a recent series of 26 cases of pancreatic carcinoma associated with acute pancreatitis. 94 Obstructive lesions at the sphincter of Oddi cited recently as a cause of pancreatitis include benign polyps, 95 and regional enteritis of the duodenum. 96

Despite the large number of known etiologic factors in acute pancreatitis, a group remains without a known cause—the idiopathic group. This group constitutes a variable proportion in the various reported series.

Diagnosis and Differential Diagnosis

Acute pancreatitis is a disease of variable intensity, and for many decades two types have been recognized: the edematous or interstitial and the hemorrhagic or necrotic. In most cases acute pancreatitis is associated with biliary tree disease or alcoholism. Acute alcoholic pancreatitis is usually seen in male patients 25 to 65 years of age who have been drinking heavily for five to ten years. There is no characteristic clinical picture in acute pancreatitis and the manifestations may vary from a bout of vague dyspepsia, with slight abdominal pain, to a fulminating collapse with shock and death. More often than not, the outstanding symptom will be

steady, severe epigastric pain, frequently radiating to the back. The most important laboratory test is an elevated serum and urinary α -amylase. In acute pancreatitis, the serum amylase is elevated within two to twelve hours of onset. If there is no rise within 24 hours, the diagnosis probably is not acute pancreatitis. The timed two-hour urinary amylase appears to be a better test because it remains elevated after the serum amylase has returned to normal, usually within two to three days. Recently, an entirely new approach to the assay of amylase has been developed using an insoluble starch labeled covalently with Remazolbrilliant blue and other dyes,97-100 Two of the methods utilizing the new starch substrates have been automated. 101,102 These new assays have been shown to be simple and rapid and the results highly reproducible. They are considered much superior to the saccharogenic or turbidimetric methods. Whereas it is known that no relation exists between the severity of pancreatic inflammation and the degree of serum amylase elevation, the fact that serum amylase might not be elevated at the time of admission is perpetually ignored, and the assays of urinary amylase are neglected. Such omissions lead to faulty evaluations of the value of amylase determinations in the differential diagnosis of acute pancreatitis.103 Recent studies have shown that the renal clearance of amylase increases in acute pancreatitis. 104-106 If acute pancreatitis is suspected and serum amylase has returned to normal, determination of the ratio of amylase clearance to simultaneously assayed creatinine clearance might have some value in supporting the diagnosis of pancreatitis. However, an increased serum lipase and two-hour urinary amylase level offer a reasonably accurate diagnosis. 107 Measurements of serum lipase activity have not been widely used because of the long incubation times involved, the instability of the substrates and the technical difficulties of the available methods. A suitable lipase assay must be rapid and must utilize a substrate that is stable, specific for lipase of pancreatic origin and is hydrolyzed to yield a product that can be easily measured. In a recently introduced rapid, automated specific assay method for lipase, monodecanoyl fluorescein is used as substrate. 108 Serum levels of lecithinase A and deoxyribonuclease activity have been found to be elevated in acute pancreatitis but determination of these factors has not gained wide use. Since trypsin and chymotrypsin are exclusively found in the pancreas, the assay of increased trypsin and chymotrypsin in serum should be specific for the distinction of acute pancreatitis from other diseases. However, chemical methods introduced for the assay of trypsin or chymotrypsin in serum are unsuitable for this purpose because of the lack of substrate specificity, lack of sensitivity and the presence of protease inhibitors.

A radioimmunoassay technique has now been developed for the determination of α -chymotrypsin in bovine and human serum, which seems to obviate the difficulties which are inherent to the chemical methods. With this technique a three-fold increase in serum chymotrypsin levels was found in patients with acute pancreatitis as compared with normal individuals. Further work is needed, however, for complete evaluation of the usefulness of this immunoassay method in the diagnosis of pancreatitis.

The differential diagnosis of acute edematous or interstitial, from hemorrhagic pancreatitis is significant in prognosis and therapy. The early detection of methemalbumin (0-24 hours) in serum of patients with hemorrhagic pancreatitis, in the presence of haptoglobin, has been suggested for this purpose. This test has been unduly discredited by recent reports. 109,110 Whereas, pancreatic enzymes, mainly trypsin, lipase and elastase can convert hemoglobin into hematin in vitro, 111 it seems that tissue enzymes might be also effective in vivo. It has been shown that methemalbumin may occur in the serum in several other conditions such as ruptured ectopic pregnancy, severe postoperative intraperitoneal hemorrhage, superior mesenteric artery thrombosis, intrahepatic hematoma, strangulation obstruction and intravascular hemolysis. However, when methemalbumin is detected within the first 24 hours in the serum or in pleural or ascites fluids in a patient with a history and enzyme studies compatible with acute pancreatitis, the diagnosis of the necrotizing form is justified. 112 Plain films of the abdomen have diagnostic value when the "sentinel loop" is present and they also contribute to the differential diagnosis of perforated peptic ulcer or other acute conditions. Additional studies are needed for the evaluation of newer techniques such as the celiac angiogram and the echogram.

Certain Clinical Features and Complications

Pain has always been considered as the main symptom in acute pancreatitis and it is usually epigastric and less often generalized over the whole abdomen. The classic "bandlike" abdominal pain is encountered in a relatively few patients. Only recently, clinicians have realized that at times acute necrotizing pancreatitis may present itself without appreciable pain. Tachycardia, sweating, nausea and vomiting are almost always present. In the course of both interstitial and hemorrhagic pancreatitis large amounts of protein-enzyme rich fluid escape around the pancreas and, in addition to hypovolemia, may cause destruction of nearby tissues.

The complicating features in acute pancreatitis are legion. Stenosing lesions of the colon¹¹³ and colonic fistulization¹¹⁴ may occur. Mesenteric fat necrosis, producing thrombosis of mesenteric vessels may result in jejunal infarction. 115,116 The urologist should include acute pancreatitis in the differential diagnosis of upper urinary tract lesions, such as changes in the left kidney producing painless hematuria.117 Rupture of the esophagus¹¹⁸ and spleen¹¹⁹ and massive intraperitoneal bleeding due to erosion of the left gastric artery¹²⁰ have been reported. Pleuropulmonary complications include basilar atelectasis, pleural effusion, pneumonitis, pulmonary embolus and infarction, empyema and respiratory difficulties due to a pseudocyst formation in the mediastinum. 121,122 A grave complication of pancreatic pseudocysts is bleeding into the cyst122,124 with subsequent rupture into the stomach, bowel or biliary tree, producing hemobilia and massive upper gastrointestinal hemorrhage. 125 Pseudocysts are relatively rare in childhood and most of the 55 reported cases are due to trauma^{126,127} or pancreatitis (mumps). Pseudocysts of the pancreas are so designated because they do not contain an epithelial lining and therefore are not true cysts. The clinical features of pancreatic pseudocysts are highly variable and frequently misinterpreted. Pseudocysts may bleed,128 erode into adjacent structures, 117,125,129,130 or rupture into the peritoneal cavity, the mediastinum¹⁸¹ or into the portal vein, 182 or they may cause obstructive jaundice or mimic neoplasm. Individualization of management is required, and is based on location and maturity of the cyst's wall for a safe internal drainage. 188,184

Perhaps the most serious and least appreciated complications of acute pancreatitis are pancreatic abscesses and lesser omentum collections. They occur in the course or following an attack of pancreatitis, and the causative organism is usually one or more Gram-negative bacteria. In a recently reported series bacterial growth was obtained in 60 of 74 cases. E. coli was seen in 16, Aerobacter aerogenes in nine and Staphylococcus aureus in eight; and more than one organism was present in 30 percent of the cases. External drainage is considered to be the standard procedure in pancreatic abscess, and without surgical intervention the outcome is usually fatal with generalized sepsis. 135,137

A number of patients with acute pancreatitis have been observed who have become acutely confused during the course of the attack. The patients become very restless, disoriented and difficult to control. The "pancreatic encephalopathy" is most probably caused by the toxic effect of circulating enzymes on the brain¹³⁸ and the pathological picture is that of widespread demyelinization and diffuse small hemorrhages in the brain. Central pontine myelinolysis associated with acute pancreatitis has also been reported.¹³⁹ Shock complicating acute pancreatitis is an ominous prognostic sign. 140 Its mechanism is at least in part a critical reduction of plasma volume, and oliguria, hyperamylasemia and metabolic acidosis are some of the features. Recent information suggests that the shock of acute pancreatitis should not be considered strictly toxic or strictly hypovolemic. French workers have introduced the term "enzymatic toxemia." Prompt plasma volume replacement favors reversal of the hemodynamic and metabolic deficits in some cases.

Nitrogen retention in acute pancreatitis indicates poor prognosis and the precise mechanism by which pancreatitis impairs renal function is unknown. It is possible that complex vascular and humoral factors are involved.

Pathogenetic Mechanisms Initiating and Propagating Pancreatic Inflammation

Despite the accumulation of a vast amount of experimental data, attemps to relate the several etiologic factors to a single pathogenetic mechanism in acute pancreatic inflammation have failed. A complete review of existing information is beyond the scope of this communication, and only pertinent recent information will be discussed. The pancreas in the average man, weighs about 70 grams (0.1 percent of body weight) and has 13 times the protein-producing capacity of the liver and reticuloendothelial system combined¹⁴¹ (4 percent of the body weight). Cannulated steer pancreas produces about 1 gram of protein per hour, and 20 percent of the dry weight of the organ constitutes enzyme-protein. With the exception of the mucosa of the gastrointestinal tract, the pancreas is the most rapidly autolyzed body tissue after death.

Normally, the pancreatic proteases and phospholipase A are stored in the exocrine cell as inactive percursors. The ergastoplasm, the Golgi complex, the zymogen granule and the cell membrane proper segregate the exportable pancreatic enzymes in the acinar cell. The same is true for the lysosomal hydrolases which co-exist with the exportable enzymes in the same cell. As an additional safety device against intracellular activation of trypsinogen (the crucial step in the mechanism of zymogen activation) two protease inhibitors are present in the exocrine cell. The interrelationship between lysosomal and exportable enzymes in the biochemical events of acute pancreatitis is not clear. A protease, probably of lysosomal origin, was recently identified in pancreatic secretion with pH optimum between 3 and 5, which is capable of activating trypsinogen into trypsin.142 Whether such a low pH can be attained in the microenvironment of the exocrine cell is a matter of speculation. The possibility of spontaneous activation of trypsinogen under certain metabolic conditions (hyperparathyroidism, for example) is also possible but requires further documentation.

Proteolytic enzymes have been recovered from pancreatic tissue during the course of experimental pancreatitis,143 and have been identified in ascites and pleural fluid in acute pancreatitis in man;7,144 phospholipase A is present in the exudate of experimental pancreatitis145 in dogs, and chymotrypsin has been detected in the serum of patients with acute pancreatitis.9 available evidence strongly suggests that the pathogenetic principle in acute pancreatitis is autodigestion, but the intricate sequence of events remains to be established. Elastase and phospholipase A appear to be significant for coagulation necrosis, vascular injury and hemorrhage. In human pancreatic tissue undergoing necrosis, lysolecithin increased and lecithin decreased, presumably as a result of phospholipase A action. 146 Elastase which has been shown to exist in zymogen form in the acinar cell,147 appears to play an important role in the destruction of elastic tissue of intrapancreatic vessel walls in hemorrhagic canine^{148,149} and human pancreatitis.¹⁵⁰ Recent evidence indicates that under certain conditions a potentiating effect might exist between trypsin and chymotrypsin on elastase as related to elastolysis.151 Trypsin appears significant in its catalytic action, by activating inactive zymogen, with resultant hydrolytic cleavage of cellular structures, elastolysis and activation of the bradykinin system. Vasoactive polypeptides may account for the glassy edema,152 and the unduly severe pain in acute pancreatitis. Pancreatic exudate was shown to contain strong kininogenase activity, 152 and could cause hypotension when given intravenously.153

Vascular factors in the pathogenesis of acute pancreatitis have received new emphasis in recent reports. In experimental pancreatitis red blood cells have been shown to obstruct lymphatics communicating with pancreatic interstitial spaces,154 with resultant intensification of the inflammatory reaction. In acute hemorrhagic pancreatitis significant reduction of both pancreatic blood flow and perfusion was found, 155 which would further enhance local ischemia.

Whereas it appears impossible at present to incriminate a single pancreatic enzyme in the pathogenesis of acute pancreatitis in man, the combined action of proteolytic and lipolytic enzymes and of vasoactive polypeptides could explain the histopathologic features and the biochemical consequences of the disease. A plausible, hypothetical, common denominator of the pathogenetic mechanism could be an increase in permeability of cellular lipoprotein membranes surrounding the exportable and lysosimal hydrolases in the acinar cell by variety of factors capable of disturbing cell metabolism and cell membrane structure. Such factors could include endotoxins, exotoxins, viral infections, low pH (acidosis), ischemia-anoxia and direct injury to the pancreas, with resultant initiation of the autocatalytic mechanism.

Metabolic Abnormalities in Acute Pancreatitis

Lipids

The appearance of lactescent serum during an attack of acute pancreatitis is well recognized and is thought to occur in from three to eight percent of cases. 156 It is noteworthy that in pancreatitis with hyperlipemia, serum amylase and lipase values may be normal and it is assumed that gross hyperlipemia interferes with the assay of these enzymes. Thus, urinary amylase determinations are of great value in the diagnosis of pancreatitis in the presence of serum lactescence. It appears that hypocalcemia or tetany or both may complicate acute pancreatitis more frequently when it is associated with hyperlipemia. In a recent series,157 seven of nineteen patients had hypocalcemia. The syndrome of hyperlipemia that follows pancreatitis is seen most commonly in alcoholics with a type IV lipoprotein electrophoretic pattern, but types I and V have also been reported.

Zieve¹⁵⁸ presented evidence in support of the thesis that hyperlipemia seen in acute pancreatitis results from the liver injury due to alcoholism and malnutrition. A number of other mechanisms have been proposed to explain the hyperlipemia due to acute pancreatitis, among them subclinical defects of lipid metabolism associated with decreased ability to clear circulating triglycerides,156 transient decrease or inhibition of lipoprotein lipase activity,159,160 and decreased availability of insulin with resultant defective clearing of dietary lipidemia.161 However, data concerning plasma insulin levels in acute pancreatitis are scanty, 162 and it would be interesting to know the insulin activity in patients with and without hyperlipemia in the acute phase of the disease. Decreased insulin reserves apparently exist in chonic pancreatitis.163-165 The difficulties in explaining the hyperlipemia of acute pancreatitis are compounded by the fact that the ingestion of large amounts of alcohol alone can result in moderate triglyceridemia. Ethanol enhances triglyceride synthesis by the liver¹⁶⁶ and the intestine, 167 might decrease lipoprotein lipase activity,168 and might increase growth hormone and 11-hydroxycorticoid levels¹⁶⁹ in plasma. High levels of plasma cortisol were also found in ten patients with acute pancreatitis and hyperlipidemia.170 It is known that, in the presence of glucocorticoids, low concentrations of growth hormone stimulate lipolysis, an effect which is said to be mediated by 3', 5'—cyclic adenosine monophosphate and is blocked by insulin. It is also known that growth hormone and glucocorticoids are involved in the mobilization of fat in animals deprived of food or insulin. Additional work is needed, however, aimed at the clarification of the role of hormonal factors in the pathogenesis of hyperlipidemia associated with acute pancreatitis and alcoholism.

Calcium

Hypocalcemia in the course of acute pancreatitis has been attributed to calcium soap formation in areas of fat necrosis. This theory does not take into account the vast calcium stores in bone, which are rapidly and readily available and can be mobilized within a few hours. Thus, Holland et al171 showed that serum calcium levels returned to normal in one hour after severe hypocalcemia was induced by infusion of sodium ethylenediamine tetraacetic acid. Large doses of parenterally administered calcium, in patients with acute pancreatitis, often do not raise serum calcium to the expected level. Two additional mechanisms have recently been proposed in order to explain the hypocalcemia of pancreatitis. Increased levels of serum glucagon known to occur during attacks of acute pancreatitis172 might produce hypocalcemia by stimulating the secretion of thyrocalcitonin with resultant inhibition of bone resorption.173 It has been shown that glucagon hypocalcemia can be induced in dogs with an intact thyroid but not after thyroidectomy, and in situ perfusion of the thyroid with glucagon resulted in a rapid fall of calcium levels. 173 However, despite this suggestive experimental evidence in dogs, glucagon infusion failed to increase serum calcitonin levels in the majority of the normal humans tested.174 On the other hand, Stern and Bell¹⁷⁵ demonstrated that in tissue culture, using embryonic rat bone labeled with ⁴⁵Ca, glucagon directly inhibits bone resorption, induced with either parathyroid hormone or dibutyryl-3'-5'-adenosine monophosphate, indicating an additional glucagon action independent of thyrocalcitonin.

Another possible mechanism for the production of hypocalcemia is the concomitant hypomagnesemia, 176 which would make bone refractory to parahormone action. 171 The incidence of hy-

pomagnesemia in acute pancreatitis should be further investigated, especially in cases of the alcoholic variety, the better to assess its role in hypocalcemia. Furthermore, serum glucagon, parahormone and thyrocalcitonin should be determined in a large number of cases of acute pancreatitis, in order to substantiate the significance of hormonal factors in this type of hypocalcemia.

Endocrine Function

During an episode of acute pancreatitis, hyperglycemia and glycosuria develop in a large proportion of patients, and some of them have permanent diabetes following the attack.¹⁷⁸ Whereas injury to the b-cells due to the pancreatic inflammation appears to be the most plausible explanation for this phenomenon, other events such as release of glucagon¹⁷⁹⁻¹⁸¹ and glucocorticoids¹⁸² might also contribute to the hyperglycemia in acute pancreatitis.

In chronic pancreatitis, the concomitant decrease of a-cell and b-cell function manifests itself by "brittle" pancreatic diabetes. This could also explain the prolonged hypoglycemic response noted in patients with chronic calcific pancreatitis183 following the intravenous administration of insulin. The effect of glucagon on pancreatic function has been studied in recent years. Administration of glucagon to dogs with pancreatic fistulae depresses the rate of flow, volume and enzyme concentration of pancreatic juice secreted by the pancreozymin/secretin-stimulated gland. 184,185 Similar observations in humans have suggested a possible regulatory role for glucagon in the control of exocrine pancreatic secretion.¹⁸⁶ The possibility has been raised of a compensatory hypersecretion of glucagon in acute pancreatitis, with resultant suppression of pancreatic and gastric secretions and inhibition of gastrointestinal motility. On the basis of these observations, it has been suggested that glucagon might have therapeutic value during an attack of pancreatitis.187

Blood Coagulation

Various coagulation abnormalities associated with acute pancreatitis have been described, but diffuse hemorrhage or widespread thrombosis occurs rarely.¹⁸⁸ The most frequent abnormalities are prolongation of the whole clotting time and

prothrombin time and a decrease of fibrinogen and factors II, VII and IX with an elevation of the antithrombin titer. The precise mechanism for these abnormalities has not been established. However, the release of pancreatic proteases (for example, trypsin, chymotrypsin and elastase) into the circulation constitutes a plausible explanation of the changes in the clotting mechanism. For instance, trypsin can activate factor X in vitro, 189 convert prothrombin into thrombin 190 and digest fibrinogen and other clotting factors. Furthermore, improvement of coagulation variables and correction of hypofibrinogenemia have been demonstrated following the administration of a protease inhibitor from beef lung (Trasylol®)* and epsilon aminocaproic acid.191 A response to these agents probably indicates an important role for proteolytic enzymes in bringing about coagulation abnormalities in acute pancreatitis. Greipp and his associates 192 observed a patient with acute pancreatitis, complicating the administration of L-asparaginase, and found very low plasma fibrinogen and abundant fibrinogen degradation products in the blood. In a recent experimental study, small doses of trypsin were injected intravenously into rabbits, with resultant rapid decrease of fibrinogen and of platelets.193 Electronmicroscopic examination of the renal glomeruli showed thrombi containing numerous platelets and fibrin, and light microscopy revealed microthrombi in several organs. Similar phenomena have been observed in experimental¹⁹⁴ and clinical hemorrhagic pancreatitis.

Further studies in vitro and in vivo are required for a better understanding of the role of various hydrolases, such as trypsin, chymotrypsin, elastase, the phospholipases and the lysosomal enzymes, in the production of coagulation abnormalities in acute pancreatitis.

Treatment

Medical Management

The treatment in acute pancreatitis includes the use of nasogastric suction, analgesics, and the intravenous infusion of colloids and crystalloids, using central venous pressure, hourly urinary output, blood pressure and hematocrit as guides. Colloid loss is replaced with human albumin and with plasma or whole blood. Seriously ill patients will require two to six units of albumin or

^{*}Not available on the commercial drug market in the United States.

plasma or whole blood the first 24 hours in order to maintain good peripheral perfusion. 195-197 In cases of hypocalcemia with tetany, intravenous infusion of calcium will alleviate tetany without necessarily restoring serum calcium levels to normal. In the presence of significant hyperglycemia or in cases of acute pancreatitis presenting as hyperosmolar coma, insulin therapy is indicated. Morphine and meperidine (pethidine) are perhaps the two drugs most commonly used for pain of biliary or pancreatic origin, but both produce a decided rise in biliary pressure. Recent evidence suggests that pantazocine is the most appropriate strong analgesic in biliary and pancreatic disease. 198 Anticholinergic preparations have been used in acute pancreatitis in order to suppress gastric and pancreatic secretion, but their effect remains unclear. 199 Their effectiveness in reducing biliary and pancreatic duct pressure is unknown. However, butylscopolamine has been found to produce a pronounced and sustained depressing effect on the morphine-induced pressure elevation in the common bile duct.²⁰⁰ The usefulness of Trasylol, the protease inhibitor obtained from beef lung, remains unproved and this type of antienzyme therapy has not gained acceptance in this country.201-203 Intra-arterial infusion directly into the celiac artery did not improve the results in a recent series.204 Trasylol does not inhibit the elastolytic activity of pancreatic elastase, and has no effect on lipase and phospholipase A. Furthermore, in vitro studies have shown that the concentration of this inhibitor sufficient to reduce the activity of a mixture of chymotrypsin and trypsin to a residual baseline reading is not capable of eliminating the potentiating effect of these enzymes on elastase activity. Recent studies with purified human trypsin have shown that this enzyme is not inhibited by soybean trypsin inhibitor and ovomucoid,205 a fact attributed to possible differences in tertiary structure of the human enzyme as compared with other trypsins. Detailed in vitro inhibition studies of all human pancreatic proteases, with naturally occurring and synthetic inhibitors are required. A recent report indicates a beneficial effect in the treatment of acute pancreatitis with a new elastase inhibitor.206 In cases of acute hemorrhagic pancreatitis the use of peritoneal lavage²⁰⁷ has been advocated as well as general supportive measures.208 For a definitive evaluation of peritoneal lavage in hemorrhagic pancreatitis²⁰⁹ without or with drainage of the thoracic duct,²¹⁰ a controlled randomized study in a sizable number of patients is highly desirable. Experimental studies in dogs have shown that agents such as low molecular weight dextran,²¹¹ fibrinolysin or heparin²¹² offer considerable protection against the development of hemorrhagic pancreatitis induced by the intraductal injection of trypsin-digested blood. The same was true for animals subjected to postganglionic sympathectomy.²¹³

The prophylactic use of antibiotics for the prevention of secondary infection in severe cases of acute pancreatitis is purely empirical at this point and no controlled studies exist in the literature concerning the effectiveness of these agents. However, the theoretical basis for their use is as follows: In acute pancreatic inflamation there is pancreatic and peripancreatic edema due to the exudation of protein-enzyme rich fluid. The peripancreatic edema may spread to the retroperineal space, the mediastinum, the mesentery and other areas. The protein rich exudate, in edematous poorly perfused tissues, is favorable to bacterial growth. Antibiotics are given with the object of providing an effective concentration in this fluid. Superimposed infection constitutes the most lethal late complication in acute pancreatitis and it is difficult to localize. Most pancreatic abscesses contain one or more Gram-negative bacillus and other bacteria. In choosing an antibiotic in acute pancreatitis, one must remember that hepatotoxicity and renal toxicity should be avoided. Every attempt should be made, of course, for identification of the causative microorganism(s) by blood and peritoneal fluid cultures followed by sensitivity tests. We have mainly used ampicillin, but chloramphenicol, gentamycin or cephalothin with or without kanamycin have been used at times, on the basis of positive blood cultures and sensitivity tests, under constant monitoring of urinary output. Whether bowel sterilization techniques could be beneficial in acute pancreatitis, as prophylaxis against infection, is unknown.

Surgical Management

Accurate diagnosis in a patient with severe abdominal pain, tachycardia and shock, especially when seen late in the course of the disease, may be difficult indeed. Earlier reports suggested a high mortality associated with laparotomy in

patients with acute pancreatitis. However, many of the deaths occurred in the terminal stage of serious illness. Recent studies indicate a low mortality with early laparotomy. 214,215 Thus, when acute pancreatitis is suspected the patient is treated vigorously for four to six hours and if the diagnosis is in doubt and the patient is deteriorating despite vigorous supportive therapy, exploratory laparotomy is indicated. The literature does not provide a clear answer to the problem of acute pancreatitis associated with biliary tree disease. Patients with gangrene or perforation of the gall bladder or acute suppurative cholangitis require operation whether or not they have hyperamylasemia or pancreatitis.⁵⁴ For patients with severe pancreatitis who respond to treatment slowly, definitive biliary tract operation should be delayed from four to six weeks after complete recovery from acute pancreatitis.²¹⁶ An upper abdominal mass suspected of being a pseudocyst, especially one that expands suddenly and gives evidence of intra-abdominal leakage, needs surgical intervention. Persistent rising jaundice necessitates surgical decompression of the biliary tree. If at laparotomy, acute hemorrhagic pancreatitis is found, adequate aspiration of the peritoneal cavity should be carried out and large sump drains should be placed around the pancreas.217 Total pancreatectomy early in necrotizing pancreatitis, as advocated by some investigators, seems overly drastic in view of the extensive local reaction present.218,219 Should a pancreatic abscess develop, with spiking fever, high leukocyte counts and evidence of intraabdominal sepsis, surgical drainage is imperative. The nonalcoholic patient presenting with severe pancreatitis and no evidece of acute cholecystitis constitutes a difficult problem. Choledocholithiasis, at the lower end of the common bile duct, is the commonest biliary lesion found in such cases. The treatment is initially conservative, with the patient observed for signs of resolution of the acute attack, and an operation is undertaken later for removal of the stones.

The mortality rate for acute hemorrhagic pancreatitis remains high, especially when renal impairment and azotemia are present. Edematous pancreatitis has a low mortality, except when delirium tremens is present concomitantly. In hemorrhagic pancreatitis the extent of necrosis, and in both the hemorrhagic and edematous forms coexisting disorders such as fatty liver,

myocardiopathy and renal disease, constitute important factors affecting the prognosis in the individual patient.

REFERENCES

- 1. Keller PJ, Cohen E, Neurath H: The proteins of bovine pancreatic juice. J Biol Chem 233:344-349, Aug 1958
- 2. Keller PJ, Allan BJ: The protein composition of human pancreatic juice. J Biol Chem 242:281-287, Jan 25, 1967
- 3. Neurath H: Mechanism of zymogen activation. Fed Proc 23:1-7, Jan-Feb 1964
- 4. Greene LJ, Rigbi M, Fackre DS: Trypsin inhibitor from bovine pancreatic juice. J Biol Chem 241:5610-5618, Dec 10, 1966
- 5. Becker V, Wilde W: de Reuck AVC, Cameron MP (Eds): In Ciba Foundation Symposium in the exocrine pancreas; normal and abnormal functions. London, Churchill, 1962, p 56
- 6. Geokas MC, Rinderknecht H, Whigham H, et al: Release of free proteolytic activity in acute bile-induced pancreatitis in the dog. Gastroenterol 56:1160, June 1969 (abstract)
- 7. Geokas MC, Olsen H, Carmack C, et al: Studies on the ascites and pleural effusion in acute pancreatitis. Gastroenterol 58:950, Jun 1970 (abstract)
- 8. Geokas MC, Rinderknecht H, Wilding P, et al: Proteolytic enzymes in human pancreatic juice in acute pancreatitis. Clin Research 16:117, Jan 1968 (abstract)
- 9. Geokas MC, Rayyis SS: Radioimmunoassay for α-chymotrypsin in human and bovine serum. Gastroenterol 60:664, Apr 1971 (abstract)
- 10. Bernard C: In Lecons de Physiologie Experimentale. Paris 2:278, 1856
- 11. Creutzfeldt W, Schmidt H: Aetiology and pathogenesis of pancreatitis. (Current concepts). Scand J Gastroent 5:Suppl 6:47-62, 1970
 12. McCutcheon AD: A fresh approach to the pathogenesis of pancreatitis. Gut 9:296-310, Jun 1968
- 13. Shapiro H, Wruble LD, Britt LG: The possible mechanism of alcohol in the production of acute pancreatitis. Surgery 60:1108-1111, Nov 1966
- 14. Pirola RC, Davis AE: Effects of ethyl alcohol on sphincteric resistance at the choledocho-duodenal junction in man. Gut 9:557-560, Oct 1968
- 15. Pirola RC, Bolin TD, Davis AE: Does alcohol cause duodenitis? Am J Dig Dis 14:239-244, Apr 1969
- 16. Pirola RC, Davis AE: Effects of intravenous alcohol on motility of the duodenum and the sphincter of Oddi: Australas Ann Med 19:24-29, Feb 1970
- 17. Kalant H: Alcohol, pancreatic secretion, and pancreatitis. Gastroenterol 56:380-384, Feb 1969
- 18. Jones SA, Steedman RA, Keller TB et al: Transduodenal sphinc-teroplasty (not sphincterotomy) for biliary and pancreatic disease. Amer J Surg 118:292-306, Aug 1969
- 19. Hiatt N, Warner NE: Serum amylase and changes in pancreatic function and structure after ligation of pancreatic ducts. Amer Surg 35:30-35, Jan 1969
- 20. Iseri OA, Lieber CS, Gottlieb LS: The ultrastructure of the fatty liver induced by prolonged ethanol ingestion. Amer J Path 48:535-545, Apr 1966
- 21. Thorpe MEC, Shorey CD: Long-term alcohol administration. Amer J Path 48:557-568, Apr 1966
- 22. Rubin E, Beattie DS, Lieber CS: Effects of ethanol on the biogenesis of mitochondrial membranes and associated mitochondrial functions. Lab Investig 23:620-627, Dec 1970
- 23. Darle N, Ekholm R, Edlund Y: Ultrastructure of the rat exocrine pancreas after long term intake of ethanol. Gastroenterology 58:62-72, Jan 1970
- 24. Sardesai VM, Orten JM: Effect of prolonged alcohol consumption in rat on pancreatic protein-synthesis. J Nutr 96:241-246, Oct 1968
 25. Orrego-Matte H, Navia E, Feres A, et al: Ethanol ingestion and incorporation of ³²P into phospholipids of pancreas in the rat. Gastroenterology 56:280-285, Feb 1969
- 26. Clayman S, Swaminathan CV, Scholefield PG: Transport and exchange diffusion of amino acids by in vitro preparations of pancreas from normal, tumor-bearing and alcohol-treated mice. Biochem Biophys Res Comm 31:553-557, May 23, 1968
- 27. Fredrickson DS, Levy RI, Lees RS: Fat transport in lipoproteins—An integrated approach to mechanisms and disorders. New Engl J Med 256:34-44, 94-103, 148-156, 215-225, 273-281, Jan-Feb 1967
- 28. Levy RI, Fredrickson DS: Diagnoses and management of hyper-lipoproteinemia. Amer J Cardiol 22:576-583, Oct 1968
- 29. Fredrickson DS, Ono K, Davis LL: Lipolytic activity of post-heparin plasma in hypertriglyceridemia. J Lip Res 4:24-33, Jan 1963
 30. Mixter CG Jr, Keynes WM, Cope O: Further experience with pancreatitis as a diagnostic clue to hyperparathyroidism. New Engl J Med 266:265-272, Feb 1962
- 31. Sharf Y, Better O, Gellei B, et al: Long-standing recurrent pancreatitis as manifested in parathyroid carcinoma. Amer J Gastroent 52:111-115, Aug 1969
- 32. Leeson PM, Fourman P: Acute pancreatitis from Vitamin-D poisoning in a patient with parathyroid deficiency. Lancet 1:1185-1186, May 28, 1966

- 33. Strum WB, Spiro HM: Chronic pancreatitis. Ann Int Med 74:264-277, Feb 1971
- 34. Herskovic T, Wakin KG, Bartholomew LG, et al: Relationship of calcium in the serum to that in the pancreatic secretions in normal and hypercalcemic states. Surgery 58:530-534, Sep 1965
- 35. Zimmerman MJ, Dreilling DA, Rosenberg IR, et al: Secretion of calcium by the cahine pancreas. Gastroenterol 52:865-870, May 1967 36. Hansky J: Calcium content of duodenal juice. Am J Dig Dis 12:725-733, Jul 1967
- 37. Kelly TR: Relationship of hyperparathyroidism to pancreatitis. Arch Surg 97:267-274, Aug 1968
- 38. Nimmo J, Finlayson ND, Smith AF, et al: The production of calcium and magnesium during pancreatic function tests in health and disease. Gut 11:163-166, Feb 1970
- 39. Strum WD, Spiro HM, Hersh T: Studies on the relationship of pancreatic juice calcium to calcific pancreatitis. Gastroenterol 58:998, Jun 1970
- 40. Gülzow M, Diwok K, Trettin HJ: Die experimentelle Kalzium-pankreatitis der Ratte. Z Ges Inn Med 20:673-678, Nov 15, 1965
- 41. Mayday GB, Pheils MT: Pancreatitis: a clinical review. Med J Aust 1:1142-1144, Jun 6, 1970
- 42. Frey CF: The operative treatment of pancreatitis. Arch Surg 98: 406-417, Apr 1969
- 43. Stobbe KC, Remine WH, Baggenstoss EJ: Pancreatic lithiasis. Surg Gynec Obstet 131:1090-1099, Dec 1970

 44. Schmidt H, Creutzfeldt W: Calciphylactic pancreatitis and pancreatitis in hyperparathyroidism. Clin Orthop 69:135-145, Mar-Apr
- 45. Davidson P, Costanza D, Swieconek JA, et al: Hereditary pancreatitis. A kindred without gross aminoaciduria. Ann Int Med 68:88-96, Jan 1968
- 46. Whitten DM, Feingold M, Eisenklam EJ: Hereditary pancreatitis. Amer J Dis Child 116:426-428, Oct 1968
 47. Lagergren C: Calcium carbonate precipitation in the pancreas, gallstones and urinary calculi. Acta Chir Scand 124:320-325, Oct 1962
- 48. Hansson K: Experimental and clinical studies in aetiologic role of bile reflux in acute pancreatitis. Acta Chir Scand Suppl 375:5-102, 1967
- 49. Schmidt H, Creutzfeldt W: The possible role of phospholipase A in the pathogenesis of acute pancreatitis. Scand J Gastroent 4:39-48,
- 50. Weiner S, Gramatica L, Voegle LD, et al: Role of the lymphatic system in the pathogenesis of inflammatory disease in the biliary tract and pancreas. Amer J Surg 119:55-61, Jan 1970
 51. Gottfries A, Nilsson S, Samuelson B, et al: Phospholipids in human hepatic bile, gall bladder bile, and plasma in cases with acute cholecystitis. Scand J Clin Lab Invest 21:168-176, 1968
- 52. Flemma RJ, Flint LM, Osterhout S, et al: Bacteriologic studies of biliary tract infection. Ann Surg 166:563-572, Oct 1967
- 53. Konok GP, Thompson AG: Pancreatic ductal mucosa as a protective barrier in the pathogenesis of pancreatitis. Amer J Surg 117:18-
- 54. Dixon JA, Hillam JD: Surgical treament of biliary tract disease associated with acute pancreatitis. Amer J Surg 120:371-375, Sep 1970 55. Gross JB, Ulrich JA, Jones JD: Urinary excretion of amino acids in a kindred with hereditary pancreatitis and aminoaciduria. Gastroenterol 47:41-48, Jul 1964
- 56. Logan A Jr. Schlicke CP, Manning GB: Familial pancreatitis. Amer J Surg 115:112-117, Jan 1968
 57. Gross JB, Jones JD: Hereditary pancreatitis: present status. Gastroenterol 58:956, Jun 1970 (abstract)
- 58. Gross JB, Gambill EE, Ulrich JA: Hereditary pancreatitis. Decription of a fifth kindred and summary of clinical features. Amer J Med 33:358-364, Sep 1962

 59. White TT, Morgan A, Hopton D: Postoperative pancreatitis. A study of seventy cases. Amer J Surg 120:132-137, Aug 1970

 60. Peterson LM, Collins JJ Jr, Wilson RE: Acute pancreatitis occuring after operation. Surg Gynec Obstet 127:23-28, Jul 1968

- 61. Cogbill CL, Song KT: Acute pancreatitis. Arch Surg 100:673-676, Jun 1970
- 62. Bardenheier JA, Kaminski DL, William VL: Pancreatitis after biliary tract surgery. Amer J Surg 116:773-776, Nov 1968
- 63. Ambromovage AM, Howard JM, Pairent FW: The twenty-four hour excretion of amylase and lipase in the urine: Correlation with the functional state and operative injury of the pancreas. Ann Surg 167:539-546, Apr 1968
- 64. Keighley MRB, Johnson AG, Stevens AE: Raised serum amylase after upper abdominal operation. Brit J Surg 56:424-427, Jun 1969
- 65. Harjola PT, Siltanen P, Appelquist P, et al: Abominal complications after open heart surgery. Ann Chir Gynaec Fenn 57:272-274, 1968
- 66. Horton EH, Murthy SK, Seal RME: Haemorrhagic necrosis of small intestine and acute pancreatitis following open-heart surgery. Thorax 23:438-445, Jul 1968
- 67. Panebianco AC, Scott SM, Dart CH Jr, et al: Acute pancreatitis following extracorporeal circulation. Ann Thorac Surg 9:562-568, Jun 1970
- 68. Johnson WC, Nabseth DC: Pancreatitis in renal transplantation. Ann Surg 171:309-314, Feb 1970
- 69. Berk JE, Smith BH, Akrawi MM: Pregnancy pancreatitis. Amer J Gastroenterol 56:216-226, Sep 1971
- 70. Tegenfeldt EG, Kirtland HB, Brown RG: Gallstones, pancreatitis and pregnancy. Amer Surg 33:88-90, Jan 1967

- 71. Montgomery WH, Miller FC: Pancreatitis and pregnancy. Obstet Gynec 35:658-664, Apr 1970
- 72. Minkowitz S, Soloway HB, Hall JE, et al: Fatal hemorrhagic pancreatitis following chlorothiazide administration in pregnancy. Obstet Gynec 24:337-342, Sep 1964
- 73. Bronsky D, Weisbery MG, Cross MC, et al: Hyperparathyroidism and acute postpartum pancreatitis with neonatal tetany in the child. Amer J Med Sci 260:160-164, Sep 1970
- 74. Riemenschneider TA, Wislon JF, Vernier RL: Glucocorticoid-induced pancreatitis in children. Pediatrics 41:426-437, Feb 1968
- 75. Musella S, Alfano C, Ciardiello A, et al: Rare case of acute pancreatitis following an overdose of cortisone. Rass Int Clin Ter 51:400-407, Apr 15, 1971
- 76. Citron BP, Halpern M, McCarron M, et al: Necrotizing angiitis sociated with drug abuse. New Engl J Med 283:1003-1011, Nov 5,
- 77. Block MB, Genant HK, Kirsner JB: Pancreatitis as an adverse reaction to salicylazosulfapyridine. New Engl J Med 282:380-382, Feb 12, 1970
- 78. Ances IG, McClain CA: Acute pancreatitis following the use of thiazide in pregnancy. South Med J 64:267-269, Mar 1971
 79. Shaw MT, Barnes CC, Madden FJ, et al: L-asparaginase and pancreatitis. Lancet 2:721, Oct 3, 1970
- 80. Pratt CB, Simone JV, Zee P, et al: Comparison of daily versus weekly L-asparaginase for the treatment of childhood acute leukemia. J Pediatr 77:474-483, Sep 1970
- 81. Boyd JF: Salmonella typhimurium colitis and pancreatitis. Lancet 2:901-902, Oct 25, 1969
 82. Veghelyi PV: Pancreatic function in scarlet fever. Pediatrics 4:94-101, Jul 1949
- 83. Koller O: Acute interstitial pancreatitis provoked by hemolytic streptococci in infected food (fish). Acta Chir Scandinav 95:358-366, 1947
- 84. Gross JB, Comfort MW: Chronic Pancreatitis. Am J Med 21:596-617, Oct 1956
- 85. Geokas MC, Olsen H, Swanson V, et al: The association of viral hepatitis and acute pancreatitis, Cal Med (in press)
- 86. Wislocki LC: Acute pancreatitis in infectious mononucleosis. New Engl J Med 275:322-323, Aug 11, 1966
- 87. Witte CL, Schanzer B: Pancreatitis due to mumps. JAMA 203:1068-1069, Mar 18, 1968
- 88. Graighead JE: Pathogenicity of the M and E variants of the encephalomyocarditis (EMC) virus. II lesions of the pancreas, parotid and lacrymal glands. Amer J Path 48:375-86, Mar 1966
- 89. Pappenheimer AM, Kunz LF, Richardson S: Passage of coxsackie virus in adult mice with production of pancreatic disease. J Exp Med 94:45-64, Jul 1951
- 90. Platt H: The occurrence of pancreatic lesions in adult mice infected with the virus of foot and mouth disease. Virology 9:484-486, Nov 1959
- 91. Glazer AM: Pancreatic necrosis in electric shock. Arch Path 39:9-10, Jan 1945
- 92. Probstein JG, Joshi RA, Blumenthal HT: Atheromatous embolization: An etiology of acute pancreatitis. Arch Surg 75:566-572, Oct 1957
- 93. Bartholomew C: Acute scorpion pancreatitis in Trinidad. Brit Med J 1:666-668, Mar 14, 1970
- 94. Gambill EE: Pancreatitis associated with pancreatic carcinoma: a study of 26 cases. Mayo Clinic Proc 46:174-177, Mar 1971
 95. Griffen WO Jr. Schaefer JW, Schindler S, et al: Ampullary obstruction by benign duodenal polyps. Arch Surg 97:444-449, Sep 1968
- 96. Legge DA: Pancreatitis as a complication of regional enteritis of the dodenum. Gastroenterology 61:834-837, Dec 1971
- 97. Rinderknecht H. Wilding P. Haverback BJ: A new method for the determination of alpha-amylase. Experientia 23:805, Oct 15,
- 98. Ceska M, Birath K, Brown B: A new and rapid method for the clinical determination of alpha-amylase activities in human serum and urine. Optimal conditions. Clin Chim Acta 26:437-444, Dec 1969
- 99. Klein B. Foreman JA, Searcy RL: New chromogenic substrate r determination of serum amylase activity. Clin Chem 16:32-38, Jan 1970
- 100. Babson AL, Tenney SA, Megraw RE: New amylase substrate ad assay procedure. Clin Chem 16:39-43, Jan 1970
- 101. Rinderknecht H, Marbach EP: A new automated method for the determination of serum alpha-amylase. Clin Chim Acta 29:107-110. Jul 1970
- 102. Hathaway JA, Hunter DT, Berrett CR: An automated method for the determination of amylase. Clin Biochem 3:217-224, 1970
 103. Adams JT, Libertino JA, Schwartz SI: Significance of an elevated serum amylase. Surgery 63:877-884, Jun 1968
 104. Blainey JD, Northam BE: Amylase excretion by the human kidney. Clin Sci 32:377-383, Jun 1967
- 105. Mulhausen R, Brown DC, Onstad G: Renal clearance of amylase during pancreatitis. Metabolism 18:669-674, Aug 1969
- 106. Levitt MD, Rapoport M, Cooperband SR: The renal clearance of amylase in renal insufficiency, acute pancreatitis, and macroamylasemia. Ann Int Med 71:919-925, Nov 1969
- 107. Seward CW: Diagnosing pancreatitis the first day: a comparison of urinary amylase and serum enzymes in pancreatic dysfunction. Southern Med J 63:286-289, Mar 1970
- 108. Fleisher M, Schwartz MK: An automated fluorometric procedure for determining serum lipase. Clin Chem 17:417-422, May 1971

- 109. Anderson MC, Toronto IR, Needleman SB, et al: Assessment of methemalbumin as a diagnostic test for acute pancreatitis. Arch Surg 98:776-780, Jun 1969
- 110. Goodhead B: Significance of methemalbuminemia in acute abdominal emergencies. Arch Surg 101:376-378, Sep 1970
- 111. Frey CF, Bradley DM, Gloze J, et al: Hematin formation and pancreatitis. J Surg Res 9:73-78, Feb 1969

 112. Geokas MC, Weissman RA, Walberg C, et al: Serum methemalbumin in acute pancreatitis. Gastroenterol 56:1161, Jun 1969 (abstract)
- 113. Mohiuddin S, Sakiyalak P, Gullick HD, et al: Stenosing lesions of the colon secondary to pancreatitis. Arch Surg 102:229-231,
- 114. Berne TV, Edmondson HA: Colonic fistulization due to pancreatitis. Amer J Surg 111:359-363, Mar 1966
- 115. Griffiths RW, Brown PW Jr: Jejunal infarction as a complication of pancreatitis. Gastroenterol 58:709-712, May 1970
- 116. Collins JJ, Peterson LM, Wilson RE: Small intestinal infarction as a complication of pancreatitis. Ann Surg 167:433-436, Mar 1968
- 117. Guerrier K, Persky L: Pancreatic disease simulating renal abnormality. Amer J Surg 120:46-49, Jul 1970
- 118. Hooper JC, Stoker TA: Rupture of the esophagus associated with acute pancreatitis. Brit J Clin Pract 24:481-484, Nov 1970
- 119. Catanzaro FP, Abiri M, Allegra S: Spontaneous rupture of the spleen and pleural effusion complicating pancreatitis. Rhode Island Med J 51:328-329, May 1968
- 120. Karmody AM, Galloway JMD: Hæmorrhagic shock in early acute pancreatitis. Brit J Surg 58:519-520, Jul 1971
- 121. Finley JW: Respiratory complications of acute pancreatitis. Amer Surg 35:591-598, Aug 1969
 122. Gee W, Foster ED, Doohen DJ: Mediastinal pancreatic pseudocyst. Ann Surg 169:420-424, Mar 1969
- 123. Dardik I, Dardik H: Patterns of hemorrhage into pancreatic pseudocysts. Amer J Surg 115:774-776, Jun 1968
- 124. Greenstein A, DeMaio EF, Nabseth DC: Acute hemorrhage associated with pancreatic pseudocysts. Surgery 69:56-62, Jan 1971
 125. Dalton WE, Lee HM, Williams GM, et al: Pancreatic pseudocyst causing hemobilia and massive gastrointestinal hemorrhage. Amer J Surg 120:106-107, Jul 1970
- 126. Wool G, Goldring D: Pseudocyst of the pancreas. Report of five cases and review of the literature. J Pediatr 70:586-594, Apr 1967
- 127. Attard J: Pseudocyst of the pancreas in a child. Brit J Surg 56:235-238, Mar 1969
- 128. Eisenbaum S, Grant RN, Cohen A: Hemorrhagic pseudocyst with pancreatitis in a ten-year-old boy. Amer Surg 36:387-388, Jun 1970
- 129. Guyer PB, Amin PH: Radiological demonstration of spontaneous rupture of pancreatic pseudocysts. Brit J Radiol 43:342-343, May 1970
- 130. Bardenheier JA, Quintero O, Barnez HB: False aneurysm in a pancreatic pseudocyst. Ann Surg 172:53-55, Jul 1970
 131. Merikas G, Stathopoulos G, Katsas A: Painless pancreatic pseudocyst ruptured into the thoracic cavity. Gastroenterol 54:101-104, Jan 1968
- 132. Zeller M, Hetz HH: Rupture of a pancreatic cyst into the portal vein. JAMA 195:869-871, Mar 7, 1966
- 133. Walker LG Jr, Stone HH, Apple DG: Pseudocysts of the pancreas: a review of 59 cases. Southern Med J 60:389-393, Apr 1967
- 134. Balfour JF: Pancreatic pseudocysts: Complications and their relation to the timing of treatment. Surg Clin N Amer 50:395-402, Apr 1970
- 135. Evans FC: Pancreatic abscess. Amer J Surg 117:537-540, Apr 1969
- 136. Bolooki H, Jaffe B, Gliedman ML: Pancreatic abscess and lesser omentum collections. Surg Gynec Obstet 126:1301-1308, Jun 1968
- 137. Kune GA: Abscesses of the pancreas. Aust N Zeal J Surg 38: 125-128. Nov 1968
- 138. Sharf B, Bental E: Pancreatic encephalopathy. J Neurol Neurosurg Psychiat 34:337-361, Jun 1971
- 139. Sherins RJ, Verity MA: Central pontine myelinolysis associated with acute hemorrhagic pancreatitis. J Neurol Neurosurg Psychiat 31: 583-588, Dec 1968
- 140. Facey FL, Weil MH, Rosoff L: Mechanism and treatment of shock associated with acute pancreatitis. Amer J Surg 111:374-381, Mar 1966
- 141. Kukral JC, Adams AP, Preston FW: Protein producing capacity of the human exocrine pancreas. Incorporation of S⁸⁵ methionine in serum and pancreatic juice protein. Ann Surg 162:63-73, Jul 1965
 142. Berndt W, Muller-Wieland K: The role of lysosomal proteases in the pathogenesis of pancreatitis. Pol Arch Med Wewnet 4:535-537, Apr-May 1970
- 143. Herva P: Experimental biliary pancreatitis in dogs. Demonstration of proteolytic activity in pancreatic tissue extracts and interference of dog serum constituents with proteolytic enzymes. Scand J Gastroenterol S: Suppl 8:9-13, 1970
- 144. Bieth J, Metais P: The simultaneous presence of trypsin and trypsin inhibitors in some pathological effusions. Clin Chim Acta 22: 639-642, Dec 1968
- 145. Hagen PO, Ofstad E, Amundsen E: Experimental acute pancreatitis in dogs. The nature of the phospholipase activity of pancreatic exudate. Scand J Gastroenterol 4:81-88, 1969

- 146. Schmidt H, Creutzfeldt W, Habermann E: Phospholipase A—ein moglichezweise entscheidender Faktor in der Pathogenese de akuten Pankreatitis. Klin Wschr 45:163-164, Feb 1, 1967
- 147. Trowbridge JO, Moon HD: Elastase in human pancreas. Immunologic and fluorescent antibody studies. Lab Investig 21:288-291, Oct 1969
- 148. Geokas MC: The role of elastase in acute pancreatitis, I. Intrapancreatic elastolytic activity in bile-induced acute pancreatitis in dogs. Arch Path 86:117-126, Aug 1968
- 149. Geokas MC: The role of elastase in acute pancreatitis. II. Intrapancreatic elastolytic activity in trypsin-induced acute pancreatitis in dogs. Arch Path 86:127-134, Aug 1968

 150. Geokas MC, Rinderknecht H, Swanson V, et al: The role of elastase in acute hemorrhagic pancreatitis in man. Lab Investig 19: 235-239. Aug 1968
- elastase in acute h 235-239, Aug 1968
- 151. Rinderknecht H, Silverman P, Geokas MC: Investigations on the standardization, assay and action of elastase. Enzymologia 40:345-
- 152. Ofstad E: Formation and destruction of plasma kinins during experimental acute hemorrhagic pancreatitis in dogs. Scand J Gastroent 5:Suppl 5:1-44, 1970
- 153. Ofstad E, Amundsen E, Hagen PO: Experimental acute pancreatitis in dogs. II. Histamine release induced by pancreatic exudate. Scand J Gastroent 4:75-79, 1969
- 154. Anderson MC, Schiller WR: Microcirculatory dynamics in normal and inflamed pancreas. Amer J Surg 115:118-127, Jan 1968
- 155. Goodhead B: Acute pancreatitis and pancreatic blood flow. Surg Gyn Obst 129:331-340, Aug 1969
- 156. Greenberger NJ, Hatch FT, Drummery GD, et al: Pancreatitis and hyperlipemia: A study of serum lipid alterations in 25 patients with acute pancreatitis. Medicine 45:161-174, Mar 1966
- 157. Cameron JL, Crisler C, Margolis S, et al: Acute pancreatitis with hyperlipemia. Surgery 70:53-61, Jul 1971
- 158. Zieve L: Relationship between acute pancreatitis and hyperlipemia. Med Clin N Amer 52:1493-1501, Nov 1968
- 159. Orvis HH, Evans JM: Serum lipids and lipid enzymes in acute pancreatitis and lipemia Clin Res Proc 5:197-198, Apr 1957
- 160. Kessler JI, Kniffen JC, Janowitz HD: Lipoprotein lipase inhibition in the hyperlipemia of acute alcoholic pancreatitis. New Engl J Med 269:943-948, Oct 1963
- 161. Bagdade JD: Diabetic lipemia complicating acute pancreatitis. Lancet 2:1041-1043, Nov 15, 1969
- 162. Rudyi RV, Chaplinsky VV: The insulin activity of the blood plasma in patients with different forms of acute pancreatitis. Klin Med (Mosk) 43:77-82, Nov 1965
- 163. Joffe BI, Bank S, Jackson WPU, et al: Insulin reserve in patients with chronic pancreatitis. Lancet 2:890-892, Oct 26, 1968
- 164. Joffe BI, Bank S, Jackson WPU: Effect of intravenous tolbuta-ide on serum insulin levels in pancreatic diabetes. Diabetes 18:499-
- 165. Rogers JB, Howard JM, Parrent FW: Serum insulin levels in patients with chronic pancreatitis. Amer J Surg 119:171-176, Feb 1970
- 166. Kudzma DJ, Schonfeld G: Alcoholic hyperlipidemia: induction by alcohol but not by carbohydrate. J Lab Clin Med 77:384-395, Mar 1971
- 167. Carter EA, Drummey GD, Isselbacher KJ: Ethanol stimulates triglyceride synthesis by the intestine. Science 174:1245-1247, Dec 11, 1971
- 168. Losowsky MS, Jones DP, Davidson CS, et al: Studies on alcoholic hyperlipemia and its mechanism. Amer J Med 35:794-803, Dec
- 169. Bellet S, Yoshimine N, DeCastro OA, et al: Effects of alcohol ingestion on growth hormone levels: their relation to 11-hydroxycorticoid levels and serum FFA. Metabolism 20:762-769, Aug 1971
 170. Geokas MC, Rayyiss SS, Chin HP, et al: Pancreatitis and hyperlipemia. Gastroenterol 56:1159, Jun 1969 (abstract)
- 171. Holland JF, Danielson E, Sahagian-Edwards A: Use of ethylenediamine tetraacetic acid in hypercalcemic patients. Proc Soc Exp Biol Med 84:359-364, Nov 1953
- 172. Paloyan E, Paloyan D, Harper P: The role of glucagon hyper-secretion in the relationship of pancreatitis and hyperparathyroidism. Surgery 62:167-173, Jul 1967
- 173. Avioli LV, Birge SJ, Scott S, et al: Role of the thyroid gland during glucagon-induced hypocalcemia in dogs. Amer J Physiol 216: 939-945, Apr 1969

 174. Tashjian AH Jr, Howard BG, Melvin KEW, et al: Immunoassay of human calcitonin. New Engl J Med 283:890-895, Oct 22, 1970
- 175. Stern P, Bell N: Effects of glucagon on serum calcium in the tand on bone resorption on tissue culture. Endocrinology 87:111-117, Jul 1970
- 176. Muldowney FP, McKenna TJ, Kyle LH, et al: Parathormone-like effect of magnesium replenishment in steatorrhea. New Engl J Med 282:61-68, Jan 8, 1970 177. Sherwood LM: Magnesium ion and parathyroid function. New Engl J Med 282:752, Mar 26, 1970
- 178. Katz LA, Spiro HM: Gastrointestinal manifestations of diabetes. New Engl J Med 275:1350-1361, Dec 15, 1966
- 179. Paloyan D, Paloyan E, Worobec R, et al: Serum glucagon levels in experimental acute pancreatitis in the dog. Surg Forum 17:348-349, 1966
- 180. Paloyan E, Paloyan E, Harper PV: Glucagon induced hypocalcemia. Metabolism 16:35-39, Jan 1967

- 181. Lawrence AM: Radioimmunoassayable glucagon levels in man: effects of starvation, hypoglycemia, and glucose administration. Proc Nat Acad Sci USA 55:316-320, Feb 1966
- 182. Geokas MC, Rayyis SS, Rinderknecht H, et al: Plasma cortisol and growth hormone levels in acute pancreatitis. Gastroenterol 56: 1160, Jun 1969 (abstract)
- 183. Joffe BI, Bank S, Marks IN: Hypoglycemia in pancreatitis Lancet 2:1038, Nov 9, 1968
- 184. Dyck WP, Rudick J, Hoexter B, et al: Influence of glucagon on exocrine pancreatic secretion. Gastroenterol 56:531-537, Mar 1969
- 185. Nakajima S, Magee DF: Inhibition of exocrine pancreatic secretion by glucagon and D-glucose given intravenously. Canad J Physiol Pharmacol 48:299-305, May 1970
- 186. Dyck WP, Texter EL Jr, Lasater JM, et al: Influence of glucagon on pancreatic secretion in man. Gastroenterol 58:532-539, Apr 1970
- 187. Knight MJ, Condon JR, Smith R: Possible use of glucagon in the treatment of pancreatitis. Brit Med J 2:440-442, May 22, 1971
- 188. Shinowara GY, Stutman LJ, Walters MI, et al: Hypercoagulability in acute pancreatitis. Amer J Surg 105:714-719, Jun 1963
- 189. Yin ET: Kinetics of human blood coagulation induced by trypsin. Thromb Diath Hæmorrh 12:307-330, Oct 15, 1964
- 190. Landman H, Markwardt F, Perlewitz J: Beeinflussung der Wirkung des trypsins auf die Blutgerinnung durch naturliche und synthetische Trypsin-und Thrombininhibitorum. Thromb Diath Hæmorrh 22:552-568, Dec 31, 1968
- 191. Wagner F: Diagnostic and therapeutic significance of the blood coagulation disorder in acute pancreatitis. Helv Chir Acta 33:118-
- 192. Greipp PR, Brown JA, Gralnick HR: Defibrination in acute pancreatitis. Ann Int Med 76:73-76, Jan 1972
- 193. Giacobino J-P, Simon GT: Experimental glomerulonephritis induced by minimal doses of trypsin. Arch Path 21:193-200, Mar 1971
- 194. Gabryelewicz A, Niewiarowski S: Activation of blood clotting and inhibition of fibrinolysis in acute pancreatitis. Thromb Diath Hæmorrh 20:409-414, Dec 31, 1968
 195. Zimberg YH: Pancreatitis: principles of management. Surg Clin N Amer 48:889-905, Aug 1968
- 196. Glieman M., Bolooki H. Rosen RG: Ravitch MM (Ed): In Current Problems in Surgery. Acute Pancreatitis. Year Book Med Publishers Inc, Chicago, 1970, P 3-52
 197. Geokas MC, Van Lancker JL, Kadell BM, et al: Acute pancreatitis. Ann Int Med 76:105-117, Jan 1972
- 198. Economou G, Ward-McQuaid JN: A cross-over comparison of the effect of morphine, pethidine, pentazocine and phenazocine on bili-ary pressure. Gut 12:218-221, Mar 1971
- 199. Bock OAA: A comment on the use of anticholinergics in the management of acute pancreatitis. South Afr Med J 42:859-860, Aug 31, 1968

- 200. Kewenter J, Kock NG: The effect of some spasmolytic drugs on the choledocho-duodenal junction in man. Scand J Gastroent 6:401-405, 1971
- 201. Baden H, Jordal K, Lund F, et al: Prophylactic and curative action of Trasylol in pancreatitis. A double-blind trial. Scand J Gastroent 4:291-295, 1969
- 202. Skinner DB, Corson JG, Nardi GL: Aprotinin therapy as prophylaxis against postoperative pancreatitis in humans. JAMA 204: 945-948, Jun 10, 1968
- 203. Hansson K, Lenninger S: Proteinase inhibitors in acute pancreatitis. Acta Chir Scand Suppl 378:103+, 1967
- 204. Vestad E, Aakhus T: Acute pancreatitis. Intra-arterial treatment with trasylol. Acta Chir Scand 136:147-151, 1970
- 205. Travis J, Roberts RC: Human trypsin. Isolation and physical-chemical characterization. Biochemistry 8:2884-2889, Jul 1969
- 206. Hansky J: The use of a peptidase inhibitor in the treatment of acute pancreatitis. Med J Aust 1:1284-1285, Jun 21, 1969
- 207. Gjessing J: Peritoneal dialysis in severe acute hemorrhagic pancreatitis. Acta Chir Scand 133:645-647, 1967
- 208. Bolooki H, Gliedman ML: Peritoneal dialysis in treatment of acute pancreatitis. Surgery 64:466-471, Aug 1968
- 209. Geokas MC, Olsen H, Barbour R, et al: Peritoneal lavage in the treatment of acute hemorrhagic pancreatitis. Gastroenterology 58: 950, Jun 1970 (abstract)
- 210. Dreiling DA: The lymphatics, pancreatic ascites and pancreatic inflammatory disease. A new therapy for pancreatitis. Amer J Gastroent 53:119-131, Feb 1970
- 211. Wright PW, Goodhead B: The value of dextrans in the treatment of experimental pancreatitis. Surgery 67:807-815, May 1970
- 212. Wright P, Goodhead B: Prevention of hemorrhagic pancreatitis with fibrinolysin or heparin. Arch Surg 100:42-46, Jan 1970
 213. Goodhead B, Wright PW: The effect of postganglionic sympathectomy on the development of hemorrhagic pancreatitis in the dog. Ann Surg 170:951-960, Dec 1969
- 214. Diaco JF, Miller LD, Copeland EM: The role of early diagnostic laparotomy in acute pancreatitis. Surg Gynec Obst 129:263-269, Aug 1969
- 215. Trapnell JE, Anderson MC: Role of early laparotomy in acute pancreatitis. Ann Surg 165:49-55, Jan 1967
- 216. Herman RE, Hertzer NR: Time of biliary surgery after acute pancreatitis due to biliary disease. Arch Surg 100:71-75, Jan 1970 217. Lawson DW, Daggett WM, Civetta JM, et al: Surgical treatment of acute necrotizing pancreatitis. Ann Surg 172:605-617, Oct 1970
- 218. Watts GT: Total pancreatectomy for fulminant pancreatitis. Lancet 2:384, Aug 24, 1963
- 219. Andersson G, Johnson SR: Treatment of acute necrotizing pancreatitis. Acta Chir Scand 134:311-313, 1968

POST-INTUBATION TRACHEAL STENOSIS

Once patients with post-intubation tracheal stenosis begin to develop symptoms, they are awfully close to obstruction. We have found in patients who are still sedentary from the disease that originally required their respiratory therapy that their airway gets down to about 5 mm before they begin to have symptoms. One of the people in our medical pulmonary unit has done some experiments with volunteers and also measured patients and found that until the airway gets down to the critical diameter of 5 to 6 mm patients at rest really don't have symptoms. This is sort of startling but it is a fact. Most of the patients we have seen have had about the same cutoff point when their condition became obvious or they became totally obstructed.

—Hermes C. Grillo, M.D., Boston
Extracted from Audio-Digest Surgery, Vol. 18, No. 10, in the Audio-Digest Foundation's subscription series of tape-recorded programs. For subscription information: 1930 Wilshire Blvd., Suite 700, Los Angeles, Ca. 90057